

Ruthenium Mediated Guanine Oxidation in Reverse Micelles

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One electron guanine (G) oxidation, a reaction linked to mutagenic events, is usually investigated in dilute solution. To model the compact environment of DNA in a cell, G oxidation has been investigated in reverse micelles (RMs). A photochemical method to generate Ru^{3+} using bis(2,2'-bipyridine)dipyridophenazine ruthenium(II) chloride, $[\text{Ru}(\text{bpy})_2\text{dppz}]\text{Cl}_2$, was used to oxidize G's in DNA. The environment of $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ in RMs was characterized by emission spectroscopy in anionic and cationic RMs. In buffer solution, $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ exhibits an emission spectrum when intercalated into DNA; no emission spectrum is observed without DNA. In anionic RMs, $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ gave identical emission spectra in the absence or presence of DNA, showing that $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ is not bound to DNA in this system, but is instead bound to the anionic RM headgroups. In cationic RMs, $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ behaves as in buffer solution, but with a 70% decrease in emission intensity. Cationic RMs were used in subsequent experiments to ensure binding of the $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ to DNA. In RMs, only $[\text{Fe}(\text{CN})_6]^{3-}$ efficiently quenches the $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ excited state to create the 3+ oxidation state. The Stern-Volmer plot of $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ and $[\text{Fe}(\text{CN})_6]^{3-}$ in RMs is curved, supporting a Poissonian distribution of reactants in the RMs (Atik, S.S, Thomas J.K, *J. Am. Chem. Soc.*, **1981**, 103, 7403). The optimal quencher concentration to generate the maximum Ru^{3+} yield was determined from the Stern-Volmer plot. Circular dichroism spectra of DNA are the same in buffer and in cationic RMs. High-resolution denaturing polyacrylamide gel electrophoresis (PAGE) is used to detect G oxidation products in buffer solution and RMs using identical concentrations of reactants. In RMs, two to three-fold longer illumination times were required to detect G oxidation products; however, similar levels of oxidation products in RMs were observed with or without $[\text{Fe}(\text{CN})_6]^{3-}$ present. This result indicates that the Ru^{3+} form of $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ does not oxidize G in RMs, but that singlet oxygen is most likely the G oxidant. By encapsulating DNA in cationic RMs, G oxidation by $[\text{Ru}(\text{bpy})_2\text{dppz}]^{2+}$ follows a different mechanism than in buffer solution. Together, these results show that the levels and mechanism of G oxidation depend intimately on the conditions under which these reactions are probed.